

Hansenia trifoliolata, a new species (Apiaceae) from Shaanxi, China

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Abstract

Hansenia trifoliolata Q.P.Jiang & X.J.He (Apiaceae), is described as new from Shaanxi Province, northwest China. The mericarp features of *H. trifoliolata* resemble *H. himalayensis* and *H. phaea* and molecular phylogenetic analyses (combining ITS and plastid genomes data) suggest that *H. trifoliolata* is closely related to the group formed by *H. oviformis* and *H. forbesii*. The new species *H. trifoliolata* has unique 3-foliate leaves and differ from other *Hansenia* species in its leaves, umbel numbers and size. A comprehensive description of *H. trifoliolata* is provided, including habitat environment and detailed morphological traits.

Keywords

Apiaceae, *Hansenia*, new species, phylogenetic analyses

Introduction

The Apiaceae is a large family with high morphological diversity, the generic and tribal delimitations within it being notoriously difficult (Shan and Sheh 1992; Plunkett and Downie 1999). Fruit characteristics have long been regarded as one of the most important sources of evidence for generic and tribal delimitation within the family (Drude 1898; Liu et al. 2003, 2007, 2009; Winter et al. 2008; Magee et al. 2010, 2011).

Notopterygium H. Boissieu (Apiaceae) was first established by Boissieu in 1903 with two species, *N. forbesii* H. Boissieu and *N. franchetii* H. Boissieu, which later appeared to be identical (Boissieu 1903). In some later studies, the genus *Notopterygium* contained six species: *N. forbesii*, *N. forrestii* H. Wolff, *N. oviforme* Shan, *N. incisum* Ting ex H. T. Chang, *N. pinnatiivolucellatum* Pu et Y. P. Wang and *N. tenuifolium* Sheh et Pu (Wolff 1930; Shan 1943; Chang 1975; Pu and Wang 1994; She and Pu 1997; Pu et al. 2000; She and Watson 2005a). The roots of *N. incisum* and *N. forbesii* are used in traditional Chinese medicine and named “Qiang Huo” (Wang et al. 1996; She and Watson 2005a; Wei et al. 2019). Although *Notopterygium* has long been treated as an endemic genus in China, Pimenov et al. (2008) merged it into the genus *Hansenia*.

Hansenia Turcz. belongs to the East Asia Clade of Apiaceae and it was first established by Turczaninow in 1844, with *H. mongolica* Turcz. as the type species (Turczaninow 1844; She and Watson 2005a; Pimenov et al. 2008; Downie et al. 2010; Pimenov 2017; Gou et al. 2020). *Hansenia* used to be treated as a monotypic genus and then some species had been transferred into this genus. Pimenov et al. (2008) transferred all the species of *Notopterygium*, except *N. tenuifolium*, to *Hansenia* through comparative morphological and molecular phylogenetic analyses and proposed five new combinations: *H. forbesii* (H. Boissieu) Pimenov & Kljuykov, *H. forrestii* (H. Wolff) Pimenov & Kljuykov, *H. oviformis* (R. H. Shan) Pimenov & Kljuykov, *H. weberbaueriana* (Fedde ex H. Wolff) Pimenov & Kljuykov and *H. pinnatiinvolutellata* (F. T. Pu & Y. P. Wang) Pimenov & Kljuykov (Pimenov et al. 2008; Pimenov 2017). Due to the lack of relevant material, *N. tenuifolium* was still retained in the genus *Notopterygium*. Subsequently, based on morphological and molecular data, Jia et al. (2019) considered that *H. pinnatiinvolutellata* was a synonym of *H. weberbaueriana*. Additionally, Tan et al. (2020), based on morphological characters and molecular data, transferred the two species of the genus *Haplosphaera* Handel-Mazzetti (Apiaceae) (She and Watson 2005b) into the genus *Hansenia* and proposed two new combinations: *H. himalayensis* (Ludlow) J.B. Tan & X.G. Ma and *H. phaea* (Handel-Mazzetti) J.B. Tan & X.G. Ma (Tan et al. 2020). Therefore, there are six species in *Hansenia* and one species in *Notopterygium* to date.

During a botanical expedition to Feng County in western Shaanxi Province in 2019, a umbelliferous species with thin stem and unusual 3-foliate leaves was collected. Species with 3-foliate leaves are rare in Apioideae and only *Trachydium trifoliatum* H. Wolff is known in China (Shan and Sheh 1992). 3-foliate leaves are commonly found in the genus *Sanicula* L. (Apiaceae) (She and Phillippe 2005), but the fruits were significantly different from *Sanicula*. After consulting relevant floras and literature, we identified that the fruit of the new species resembles *H. himalayensis* and *H. phaea* and further molecular phylogenetic analyses supported our conclusion. Based on careful morphological and molecular analyses, we identified it as a new species of *Hansenia*.

Materials and methods

DNA extraction and sequencing

Fresh leaves of *Hansenia trifoliolata* were collected from wild plants, desiccated and stored in silica gel. The herbarium specimens were stored in the Herbarium, College of Life Sciences, Sichuan University (SZ). Specimen voucher details were provided in Table 1 and Suppl. material 1: Fig. S2. Total genomic DNA was extracted from the stored dry leaves, using a CWBIO plant genomic DNA extraction kit (CWBIO, Beijing, China), following the manufacturer's protocols. PCR-amplification of the complete ITS region used the primers of ITS4 (5'-TCC TCCGCT TAT TGA TAT GC-3') and ITS5 (5'-GGA AGTAAA AGT CGT AAC AAG G-3'; White et al. 1990). PCR amplification was undertaken in a 30 µl volume reaction, containing 3 µl plant total DNA, 1.5 µl of each forward primer and reverse primer, 10 µl ddH₂O and 15 µl 2×Taq MasterMix (CWBIO, Beijing, China). The PCR amplification of the nrITS region had an initial denaturation for 4 min at 94 °C, followed by 30 cycles of 45 s at 94 °C, 45 s at 53 °C and 60 s at 72 °C, then a final extension of 10 min at 72 °C. All PCR products were sent to Sangon (Shanghai, China) for sequencing after being examined using a 1.5% (w/v) agarose TAE gel. The DNA sequences of nrITS were applied for phylogenetic analyses and detailed information as outlined in Table 1.

Plastid genome sequencing, assembly and annotation

We sequenced, assembled and annotated the plastid genome of *Hansenia trifoliolata*, then compared it with other species of *Hansenia*. The processes of plastid genome sequencing, assembly and annotation were performed as follows.

The Illumina Novaseq 6000 platform (Illumina, San Diego, CA, USA) at Novogene (Beijing, China) was used to sequence the resultant DNA with Novaseq 150 sequencing strategy. The remaining clean data were assembled using NOVOPlasty 2.7.1 (Dierckxsens et al. 2017) with the default K-mer value 39 and rbcL of *H. oviformis* (GenBank accession No.: MF787597.1) being used as seed input. Preliminary genome annotation was conducted using PGA (Qu et al. 2019), with manual modifications for uncertain genes and uncertain start and stop codons, based on comparison with other related plastid genomes, using Geneious R11 soft (Kearse et al. 2012). Protein-coding sequence (CDS) was extracted from the plastid genome using the PhyloSuite programme (Zhang et al. 2020). The plastid genome of *H. trifoliolata* was submitted to GenBank and the accession number was listed in Table 1.

Table 1. Voucher details and GenBank accession number of *Hansenia trifoliolata*.

Taxa (Species number)	Voucher	Locality	Genbank number	
			Plastid genome	ITS
<i>Hansenia trifoliolata</i>	JQP19082004	Feng County, Shannxi Province	OM281945	OM800961 OM800962

Phylogenetic analyses

We used MEGA7 (Kumar et al. 2016) to align DNA sequences with manual adjustment to improve the accuracy of sequence alignment. Phylogenetic analyses were undertaken applying Maximum Likelihood (ML) and Bayesian Inference (BI) analyses. Based on the Akaike Information Criterion (AIC) implemented in MrModelTest version 2.2 (Nylander 2004), the best-fit nucleotide substitution models for the ITS sequences (GTR+G) and protein-coding sequences (GTR+G+I) were determined, respectively. ML analyses were undertaken using RAxML v.8.2.4 (Stamatakis 2014) with the best-fit model and 1000 bootstrap replicates. BI analyses were conducted with MrBayes version 3.2 (Ronquist et al. 2012). Four simultaneous runs were performed using Markov Chain Monte Carlo (MCMC) simulations for 10 million generations, starting from a random tree and sampling one tree every 1000 generations. The first 20% of obtained trees were discarded as burn-in and the remaining were used to calculate a majority-rule consensus topology and posterior probability (PP) values.

Results

Morphological study

We collected several specimens of *H. trifoliolata* from Feng County, Shannxi Province and the type locality at an elevation of 2300–2500 m (Fig. 1). After field observation, we investigated the fruit morphological characteristics of *H. trifoliolata* (Fig. 2), including fruit shape and size, ribs, vittae and endosperm which are highly similar to *H. himalayensis* and *H. phaea*. We compared the morphological characteristics of *H. trifoliolata* with the related species (*H. himalayensis*, *H. phaea*, *H. oviformis*, *H. forbesii* and *H. forestii*), including life form, leaves, umbel rays and fruits (Table 2).

Phylogenetic analyses

The phylogenetic analysis result, based on ITS data, is shown in Fig. 3. The details of the ITS dataset that we sequenced for phylogenetic analysis are listed in Table 1. The phylogenetic trees derived from BI and ML analyses were topologically consistent. Thus, only the BI tree is shown in Fig. 3, with bootstrap support values obtained from ML analyses. The phylogenetic tree showed that *H. trifoliolata* was sister to *H. oviformis*, with strong support (Bayesian inference posterior probability, BI = 1.00; maximum parsimony bootstrap, ML = 96%). Additionally, *H. trifoliolata* and other *Hansenia* species formed a monophyletic group with the support very close to maximum (BI = 1.00; ML = 99%).

The result of the phylogenetic analysis, based on the plastid genome data, is shown in Fig. 4. The plastid genome GenBank number of *H. trifoliolata* is listed in Table 1.

The phylogenetic trees derived from BI and ML analyses were topologically consistent. Therefore, only the BI tree is shown in Fig. 4, with bootstrap support values obtained from ML analyses. The phylogenetic tree showed that *H. trifoliolata* clustered with the communities of *H. oviformis* and *H. forbesii* (BI = 1.00; ML = 68%). This is the same as the ITS tree, with *H. trifoliolata* and other *Hansenia* species forming a monophyletic group with maximum support (BI = 1.00; ML = 100%).

Table 2. Diagnostic morphological characters of *Hansenia trifoliolata* and related species.

Character	Taxon					
	<i>H. trifoliolata</i>	<i>H. himalayensis</i>	<i>H. phaea</i>	<i>H. oviformis</i>	<i>H. forbesii</i>	<i>H. forestii</i>
Live form	monocarpic	polycarpic	polycarpic	monocarpic	polycarpic	monocarpic
Plant height (cm)	60–90	80–120	55–90	40–60	80–180	50–100
Leaf in outline (basal)	blade broad-triangular, 3-foliolate	blade ovate-triangular, 3-pinnate	blade broad-triangular or triangular-ovate, ternate-1–2-pinnate	broadly triangular 2-pinnate	oviform 3-pinnate	broadly triangular, 2-pinnate
Median leaflets (pinnae) (basal)	cuneate-obovate or rhombic, base cuneate, with irregularly doubly serrate, apex obtuse	pinnatifid, pinnae 3–6 pairs, triangular or narrowly ovate-triangular, ultimate segments, mucronate, acute-dentate	ovate or obovate, 3-parted, base cuneate; with irregularly doubly serrate or serrate, apex obtuse	(broadly) obovate to almost round, base cuneate, margins serrulate, apex obtuse	broadly lanceolate to oviform-lanceolate, base obtuse or cuneate, margins serrate	oviform to lanceolate, base cuneate, margins irregular or sharply serrate
Lateral leaflets (pinnae) (basal)	oblique-ovate, base oblique, often shallowly or deeply uneven 2-parted or not divided; irregularly doubly serrate, apex obtuse	pinnatifid, pinnules 3–4 pairs, ultimate segments mucronate, acute-dentate	ovate to ovate-lanceolate, base oblique; with irregularly doubly serrate or serrate, apex obtuse	ovate or elliptic, base truncate; margins serrulate, apex obtuse	broadly lanceolate to oviform-lanceolate, base obtuse or cuneate; margins serrate	oviform to lanceolate, base cuneate, base oblique; margins irregular or sharply serrate
Umbels	compound umbel, 3–7-rayed, unequal	compound umbel, 2–6-rayed, subglobose, unequal	Subglobose	compound umbel, 5–9-rayed, rays very unequal	compound umbel, 11–20-rayed, rays ± equal	compound umbel, 6–9-rayed, unequal
Calyx teeth	ovate-triangular, 0.3–0.5 mm	inconspicuous, triangular, ca. 0.1 mm	ovate-triangular, 0.4 × 0.5 mm	short, triangular, ca. 0.4 mm	short, lanceolate, ca. 0.5 mm	ovate-lanceolate, 0.3–0.6 mm
Fruit	ovoid-oblong or long-ellipsoid, 4–6 mm × 1.4–2.1 mm; constricted at the commissure	ovoid-oblong or long-ellipsoid, 6–7 mm × 1.5–2 mm, slightly constricted at the commissure	ovoid-oblong, 4–5 mm × 2–2.5 mm; no constricted at the commissure	globose, 4–5 × 2–3 mm; no constricted at the commissure	oblong-ellipsoid, ca. 5 × 4 mm; no constricted at the commissure	subglobose, ca. 3–3.5 × 2.5–3 mm; no constricted at the commissure
Stylopodium	conic	low-conic	depressed	flat	conic	depressed
Mericarp ribs	± equal, prominent to narrow-winged	± equal, conspicuous, narrowly winged	± equal, narrow-winged	± equal, broadly winged	± equal, winged	± equal, winged
Endosperm (at commissural side)	concave	deeply concave	concave	slightly concave	broadly and not deeply concave	concave
Vittae in dorsal furrows	3 (4)	3	3	1–2	2–4	3
Vittae in commissure	2–5	6	4–6	4	4–5	4–6

Discussion

The fruits of *H. trifoliolata* were similar to *H. himalayensis* and *H. phaea* in fruit shape and size, mericarp ribs and both vittae in dorsal furrows and in the commissure. Additionally, the endosperm (at the commissural side), slightly or deeply concave, was common in *Hansenia* (Pimenov et al. 2008; Tan et al. 2020). The fruit shape of *Hansenia* can be divided into two groups by the shape and ribs: fruits oblong-ellipsoid, subglobose or globose, all ribs winged or broadly winged ribs (including *H. forrestii*, *H. forbesii*, *H. mongolica*, *H. oviformis* and *H. weberbaueriana*); fruits obovoid-oblong or long-ellipsoid, ribs prominent to narrowly winged (including *H. trifoliolata*, *H. himalayensis* and *H. phaea*). All species' fruit ribs are 5 and ± equal, except for *H. weberbaueriana* where the ribs are 3–5, equal or a little unequal (Jia et al. 2019). Moreover, there is a constriction at the commissure in *H. trifoliolata*, with a similar phenomenon being found in *H. himalayensis* that has a slight constriction at the commissure (She and Watson 2005a; Tan et al. 2020).

The life form of *H. trifoliolata* is monocarpic, which is uncommon in *Hansenia*, except for *H. forrestii* which seems to be similar (Pimenov et al. 2008). Through observation of the specimens of *H. oviformis*, we believed that *H. oviformis* is monocarpic. The leaves of *Hansenia* species are often 1–3-pinnate, leaflets pinnatifid (including *H. mongolica*, *H. himalayensis* and *H. weberbaueriana*) or leaflets not pinnatifid (including *H. forrestii*, *H. forbesii*, *H. oviformis*, *H. phaea* and *H. trifoliolata*). The stylopodium shape in the genus is continuous, from depressed to flat, to low-conic and conic. All species of *Hansenia* display compound umbels, except for *H. phaea* and the number of rays are either below ten (including *H. trifoliolata*, *H. himalayensis*, *H. oviformis* and *H. forrestii*) or ten to twenty (including *H. forbesii*, *H. mongolica* and *H. weberbaueriana*) (Pimenov et al. 2008; Tan et al. 2020).

In our phylogenetic analyses, *H. trifoliolata* and other *Hansenia* species formed a monophyletic group in both ITS and plastid trees with very strong support (ITS trees: BI = 1.00, ML = 99%; plastid trees: BI = 1.00, ML = 100%). Though the position of *H. trifoliolata* within *Hansenia* had a slight difference between ITS trees and plastid trees (ITS trees: *H. trifoliolata* was sister to *H. oviformis*, then clusters with *H. forbesii*; plastid trees: *H. trifoliolata* clustered with the communities of *H. oviformis* and *H. forbesii*), there is no doubt that *H. trifoliolata* is a member of the genus *Hansenia*.

H. trifoliolata overlaps in its distribution with *H. forbesii* and *H. weberbaueriana* in the western Shaanxi Province and south-eastern Gansu Province.

The molecular data and morphological evidence strongly support the circumscription of *H. trifoliolata* as a new species belonging to *Hansenia*.

Key to the species of *Hansenia*

- | | | |
|----|--|---|
| 1a | Fruit oblong-ellipsoid, subglobose or globose or elliptic, all ribs winged or broadly winged, wings equal or unequal | 2 |
| 2a | Rays below ten, unequal | 3 |

- 3a Ultimate leaf segments ovate-lanceolate, 2.5–8 cm; bracteoles linear, shorter than flowers *H. forrestii*
- 3b Ultimate leaf segments ovate, 1.5–3.5 cm; bracteoles filiform, longer than flowers *H. oviformis*
- 2b Rays ten to twenty, ± equal 4
- 4a Leaves pinnatisect, leaflets pinnatifid 5
- 5a Bracteoles linear or pinnatifid, fruit ribs 3–5, ultimate leaf segments oblong, margin pinnatifid or variously laciniate-dentate *H. weberbaueriana*
- 5b Bracteoles linear, ribs 5, ultimate leaf segments broadly ovate to oblong, at the margin toothed, teeth obtuse *H. mongolica*
- 4b Leaves pinnate, leaflets not pinnatifid, ultimate leaf segments ovate to oblong-ovate, margin entire or coarsely toothed *H. forbesii*
- 1b Fruit ovoid-oblong or long-ellipsoid, ribs prominent to narrowly winged 6
- 6a Basal leaves and cauline leaves 3-foliolate, umbels 2–5 cm across, rays unequal *H. trifoliolata*
- 6b Basal leaves ternate-1–3-pinnate, flowers densely crowded into a compact, globose heads 7
- 7a Basal leaves ternate-1–2-pinnate; petals obovate, apex narrowly inflexed *H. phaea*
- 7b Basal leaves 3-pinnate; petals broad-ovate, spoon-shaped apex acute *H. himalayensis*

Description of the new species

Hansenia trifoliolata Q.P.Jiang & X.J.He, sp. nov.

urn:lsid:ipni.org:names:77307988-1

Figs 1, 5; Suppl. material 1: Fig. S2

Diagnostic characters. Monocarpic. Root cylindrical, branched or partial rhizomes. Leaves 3-foliolate. Umbels 2–5 cm across, rays 3–7, unequal. Stylopodium conical. Fruits are ovoid-oblong or long-ellipsoid, have 5 ribs, ribs prominent to narrowly winged and endosperm (at commissural side) concave. It is clearly distinguished from *H. phaea* and *H. himalayensis* in leaves (ternate-1–2-pinnate and 3-pinnate vs. 3-foliolate). Compared to other *Hansenia* species (i.e. *H. forrestii*, *H. oviformis* and *H. forbesii*), *H. trifoliolata* also shows distinctive morphological characters, especially in fruits characters (shape and ribs) and leaves (3-foliolate is unique in *Hansenia*).

Type. CHINA, Shaanxi Province: Tongtianhe National Forest Park, Feng County, elevation 2430 m a.s.l., 34°14'N, 106°33'E, 28 Sep 2021, Q. P. Jiang, JQP21092801, fruiting (Holotype: SZ).

Description. Biennial, herb, 60–90 cm high. Root cylindrical, branched or partial rhizomes. Stem purplish-green, thinly ribbed, glabrous, thin. Leaves 3-foliolate, green, blade broad-triangular, irregularly doubly serrate, teeth mucronate; central leaflets cuneate-obovate or rhombic, 4–6 × 2–3.5 cm, with irregularly doubly serrate, base

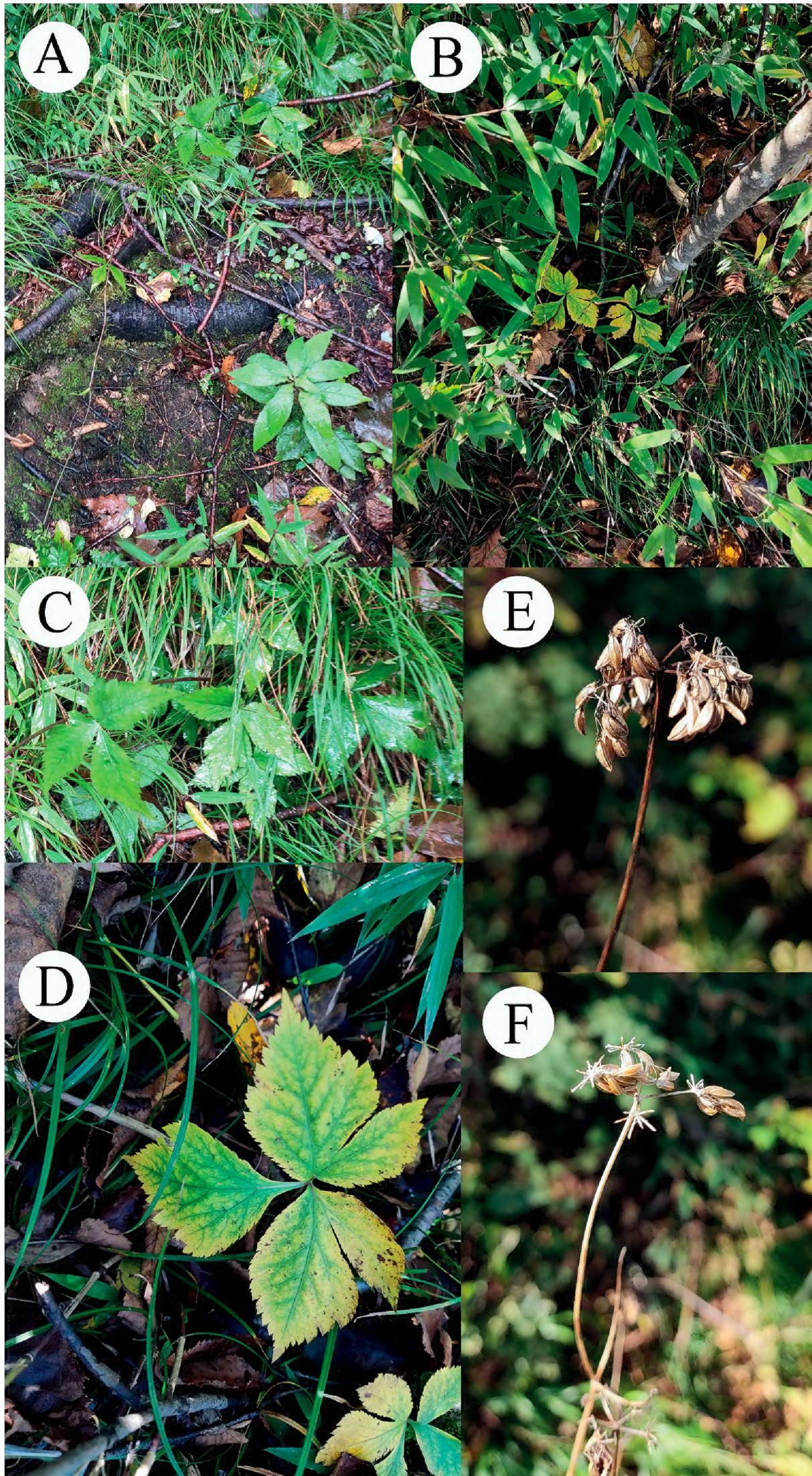


Figure 1. *Hansenia trifoliolata* in the field **A, B** habitat **C** cauline leaves **D** basal leaves **E, F** umbels and fruits.

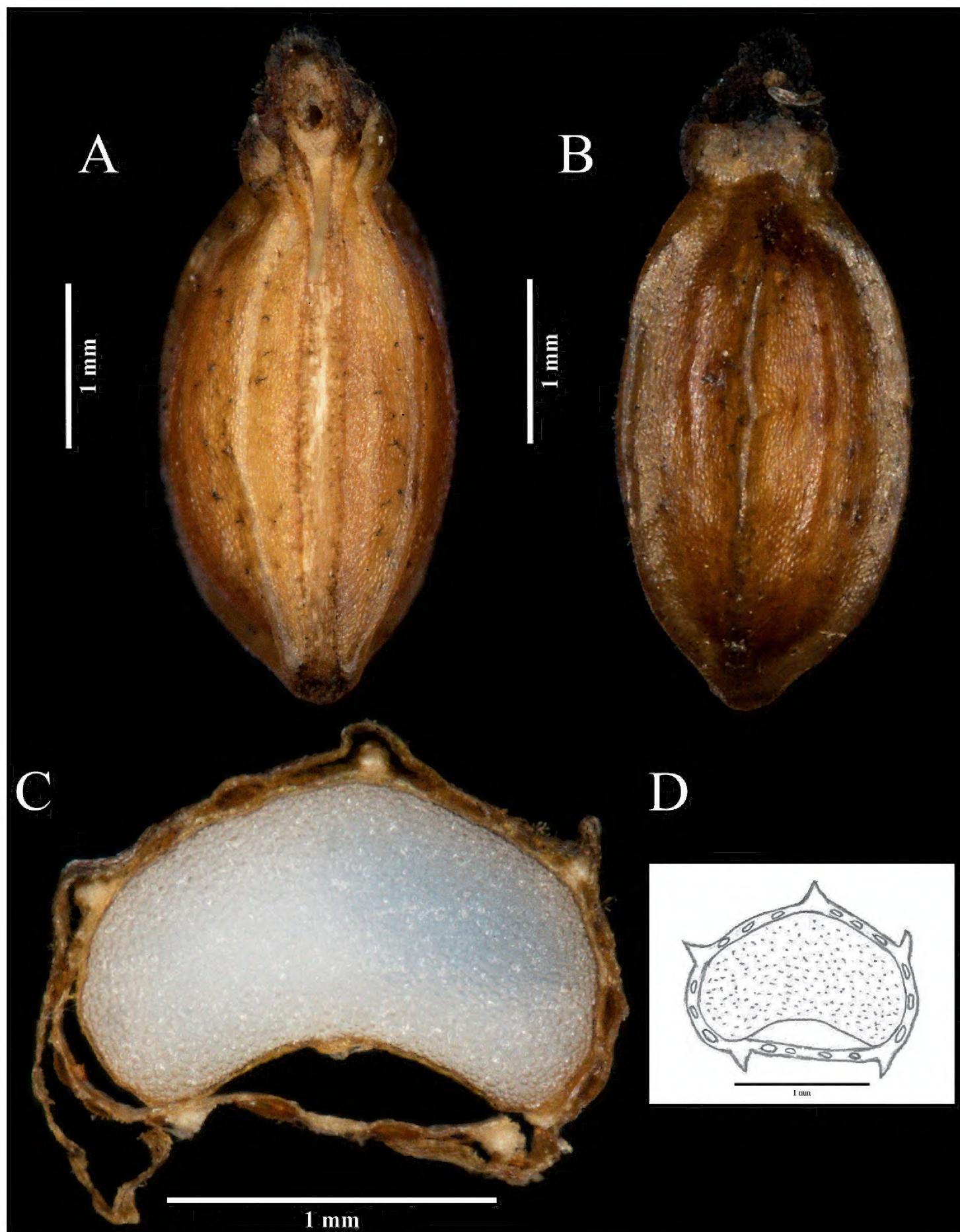


Figure 2. Fruit characters of *Hansenia trifoliolata* **A** commissural side of fruit **B** dorsal view of fruit **C** cross-section of fruit **D** the illustration of the fruit in transverse section. Voucher: JQP21092801.

cuneate; lateral leaflets oblique-ovate, base oblique, often shallowly or deeply uneven 2-parted or not divided, $2-5 \times 3.5-6.5$ cm. Basal petioles 15–20 cm, petioles shorten upwards; sheaths narrow-oblong, glabrous, with margin irregularly coarse-cuspidate-

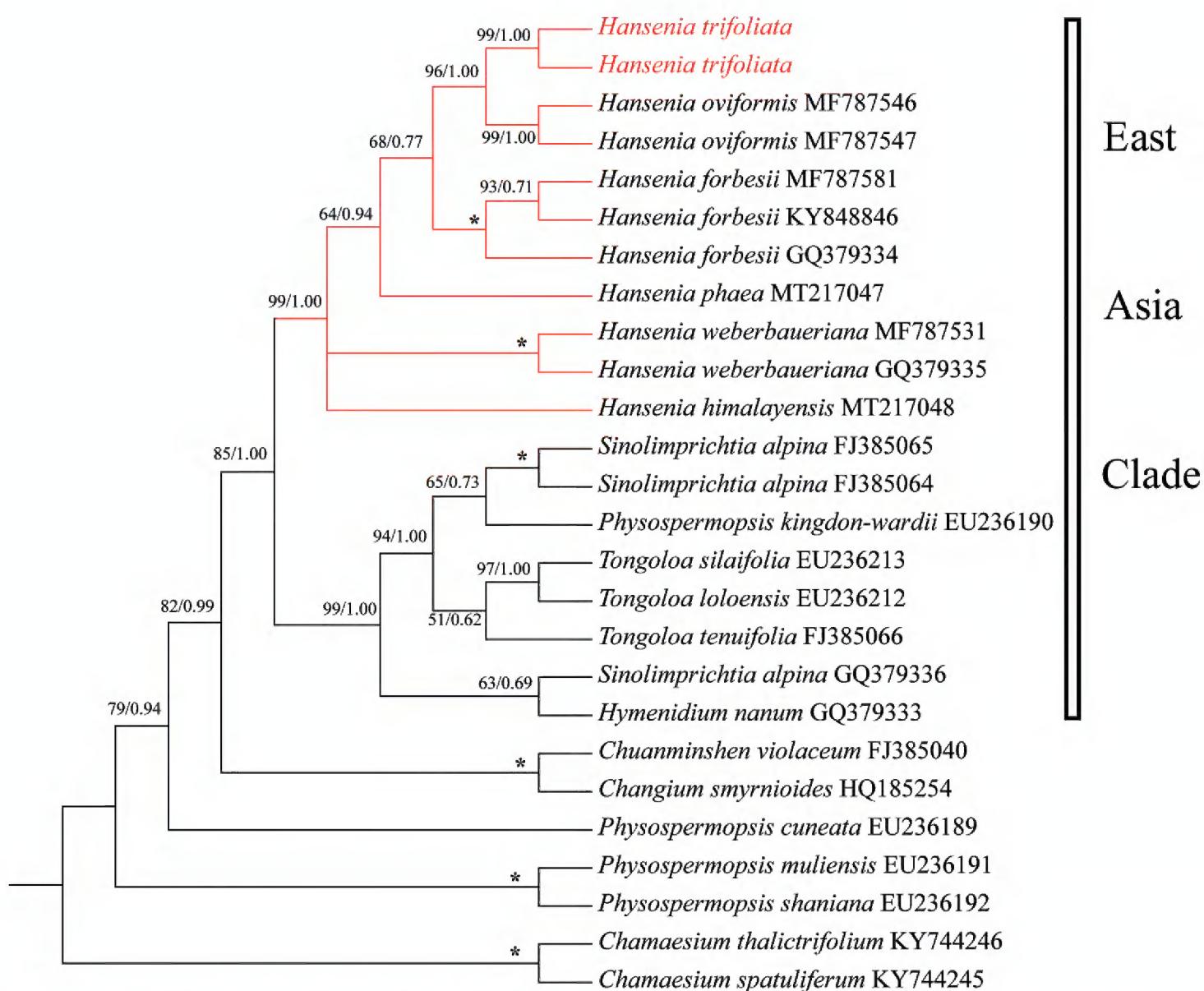


Figure 3. Bayesian 50% majority-rule consensus tree of *Hansenia trifoliolata*, other species of *Hansenia* and related species inferred from ITS sequences using a GTR+G nucleotide substitution model. The tree is rooted with two species of *Chamaesium*. Maximum Likelihood bootstrap support (ML BS) and Bayesian posterior probabilities (BI PP) are presented at the nodes, * representing the best support (100%). The ITS sequences obtained from NCBI exhibited the GenBank number adjacent to the species names.

serrate. Umbels 20–50 mm across; peduncles 5–20 mm long, glabrous; bracts 0 to 2, linear; rays 3 to 7, 5–25 mm long, glabrous; bracteoles 2 to 7, linear, 3–8 mm long; raylets 5 to 11, 1–3 mm long. Flowers unknown; calyx teeth ovate-triangular, 0.3–0.5 mm; petals unknown; stylopodium conical. Fruit ovoid-oblong or long-ellipsoid, 1.4–2.1 × 4–6 mm; mericarps 5-ribbed, ribs prominent to narrow-winged; vittae 3 (4) in each furrow, 2–5 on commissure; endosperm (at commissural side) concave, commissure width 0.8–1.35 mm.

Etymology. The specific epithet refers to the distinctive 3-foliate leaves.

Phenology. Flowering from July to August, and fruiting from August to September.

Distribution, habitat and ecology. At present, this new species has only been found in the type locality in Tongtianhe National Forest Park, Feng County, Shaanxi

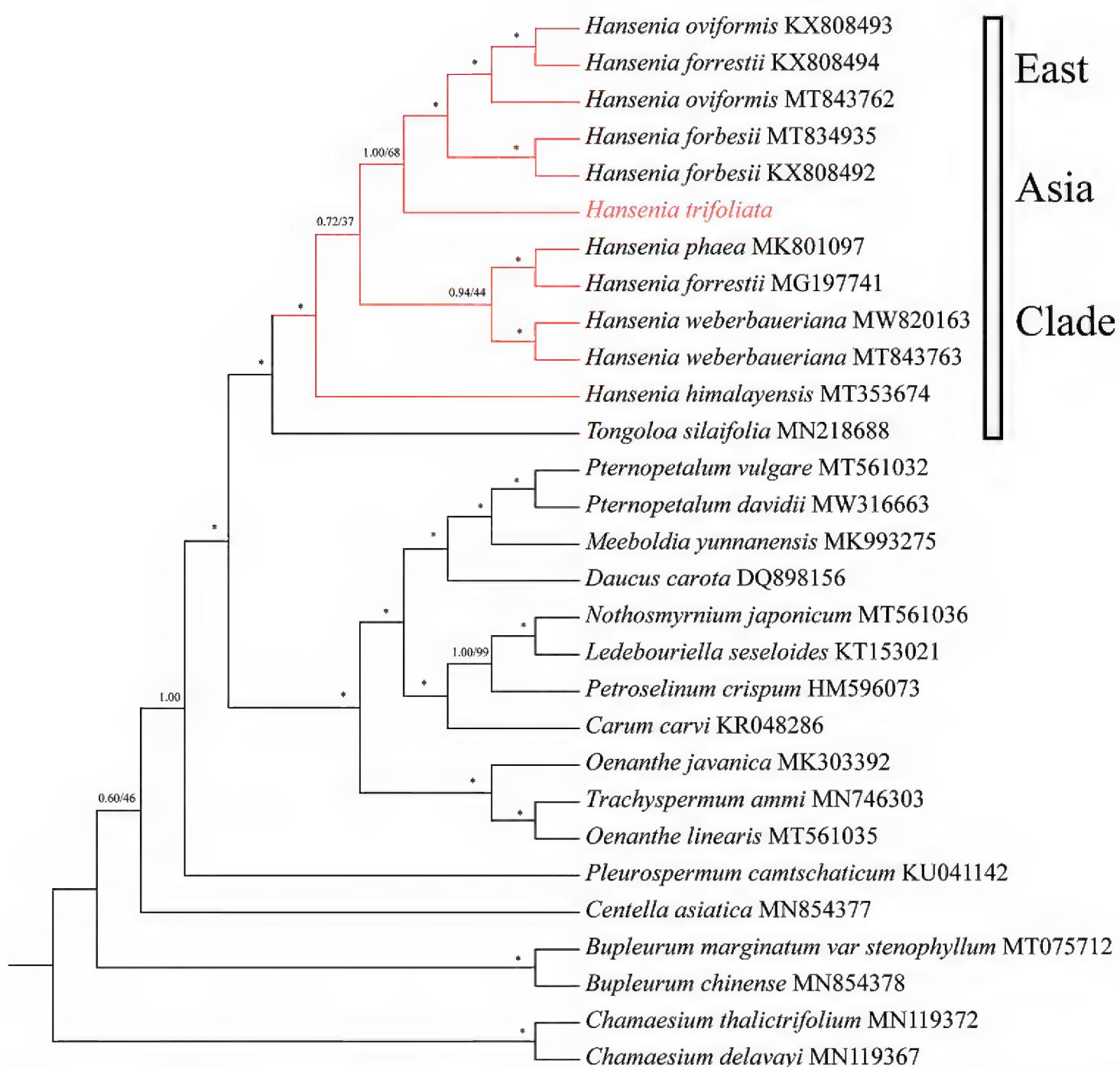


Figure 4. Bayesian 50% majority-rule consensus tree of *Hansenia trifoliolata*, other species of *Hansenia* and related species inferred from protein-coding genes of plastid genomes using a GTR+G+I nucleotide substitution model. The tree is rooted with two species of *Chamaesium*. Maximum Likelihood bootstrap support (ML BS) and Bayesian posterior probabilities (BI PP) are presented at the nodes,* representing the best support (100%). The plastid genome sequences obtained from NCBI exhibited the GenBank number adjacent to the species names.

Province, China. According to the growing environment, we speculate it may inhabit forests at an elevation of 2300 m to 2500 m in western Shaanxi Province and south-eastern Gansu Province. This new species grows in humid environments under the forests.

Additional specimens examined (paratypes). CHINA: Shaanxi Province, Baoji City, Feng County, Tongtianhe National Forest Park, elevation 2430 m a.s.l., 34°14'N, 106°33'E, 20 Aug 2019, Q. P. Jiang and X. Y. Zhang, JQP19082004 (photo SZ !).



Figure 5. Holotype of *Hansenia trifoliolata*, fruiting. Vouchers: JQP21092801.

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References

- Boissieu HD (1903) Les Ombellifères de Chine d'après les collections du Muséum d'Histoire naturelle de Paris. Bulletin de l'Herbier Boissier 3: 837–856.
- Chang HT (1975) Revision of *Notopterygium* (Umbelliferae). Journal of Systematics and Evolution 13(3): 83–87. <https://www.jse.ac.cn/EN/Y1975/V13/I3/83>
- Dierckxsens N, Patrick M, Guillaume S (2017) NOVOPlasty: De novo assembly of organelle genomes from whole genome data. Nucleic Acids Research 45: e18. <https://doi.org/10.1093/nar/gkw955>
- Downie SR, Spalik K, Katz-Downie DS, Reduron J-P (2010) Major clades within Apiaceae subfamily Apioideae as inferred by phylogenetic analysis of nrDNA ITS sequences. Plant Diversity and Evolution 128(1–2): 111–136. <https://doi.org/10.1127/1869-6155/2010/0128-0005>
- Drude O (1898) Umbelliferae. In: Engler A, Prantl K (Eds) Die natürlichen Pflanzenfamilien, div. 3, vol. 8. Wilhelm Engelmann, Leipzig, 63–150.
- Gou W, Jia SB, Price M, Guo XL, Zhou SD, He XJ (2020) Complete Plastid Genome Sequencing of Eight Species from *Hansenia*, *Haplosphaera* and *Sinodielsia* (Apiaceae): Comparative Analyses and Phylogenetic Implications. Plants 9(11): 1523. <https://doi.org/10.3390/plants9111523>
- Jia SB, Guo XL, Zhou SD, He XJ (2019) *Hansenia pinnatiinvolutellata* is conspecific with *H. weberbaueriana* (Apiaceae) based on morphology and molecular data. Phytotaxa 418(2): 203–210. <https://doi.org/10.11646/phytotaxa.418.2.5>
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A (2012) Geneious Basic: An integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics 28(12): 1647–1649. <https://doi.org/10.1093/bioinformatics/bts199>
- Kumar S, Stecher G, Tamura K (2016) Mega7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. Molecular Biology and Evolution 33(7): 1870–1874. <https://doi.org/10.1093/molbev/msw054>
- Liu M, van Wyk B-E, Tilney PM (2003) The taxonomic value of fruit structure in the subfamily Saniculoideae and related African genera (Apiaceae). Taxon 52(2): 261–270. <https://doi.org/10.2307/3647394>
- Liu M, van Wyk B-E, Tilney PM (2007) Irregular vittae and druse crystals in *Steganotaenia* fruits support a taxonomic affinity with the subfamily Saniculoideae (Apiaceae). South African Journal of Botany 73(2): 252–255. <https://doi.org/10.1016/j.sajb.2006.10.003>
- Liu M, Van Wyk B-E, Tilney PM, Plunkett GM, Lowry PP II (2009) Evidence from fruit structure supports in general the circumscription of Apiaceae subfamily Azorelloideae. Plant Systematics and Evolution 280(1–2): 1–13. <https://doi.org/10.1007/s00606-009-0160-1>
- Magee AR, Calviño CI, Liu M, Downie SR, Tilney PM, Van Wyk B-E (2010) New tribal delimitations for the early diverging lineages of Apiaceae subfamily Apioideae. Taxon 59(2): 567–580. <https://doi.org/10.1002/tax.592021>
- Magee AR, van Wyk B-E, Tilney PM, Vessio N (2011) A Taxonomic Revision of the *Annesorhiza triternata* Group (Apiaceae, Apioideae): The Transfer of *Peucedanum triternatum* and

- P. filicaule* and the Description of Five New Species. Systematic Botany 36(2): 508–519. <https://doi.org/10.1600/036364411X569697>
- Nylander JAA (2004) MrModeltest, a program to evaluate the fit of several models of evolution to a given data and unrooted tree (version 2.2). – Evolutionary Biology Centre, Uppsala University, Sweden.
- Pimenov MG (2017) Updated checklist of Chinese Umbelliferae: Nomenclature, synonymy, typification, distribution. Turczaninowia 20(2): 106–239. <https://doi.org/10.14258/turczaninowia.20.2.9>
- Pimenov MG, Kljuykov EV, Ostroumova TA (2008) Reduction of *Notopterygium* to *Hansenia* (Umbelliferae). Willdenowia 38(1): 155–172. <https://doi.org/10.3372/wi.38.38110>
- Plunkett GM, Downie SR (1999) Major lineages within apiaceae subfamily apioideae: A comparison of chloroplast restriction site and dna sequence data. American Journal of Botany 86(7): 1014–1026. <https://doi.org/10.2307/2656619>
- Pu FD, Wang YP (1994) A new species of *Notopterygium* (Umbelliferae) from Sichuan. Journal of Sichuan University (Natural Science Edition) 31(3): 386–388.
- Pu FD, Wang PL, Zhen ZH, Wang YP (2000) A reclassification of *Notopterygium* Boissieu (Umbelliferae). Journal of Systematics and Evolution 38(5): 430–436.
- Qu XJ, Moore MJ, Li DZ, Yi TS (2019) PGA: A software package for rapid, accurate, and flexible batch annotation of plastomes. Plant Methods 15(1): 50. <https://doi.org/10.1186/s13007-019-0435-7>
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Hohna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3): 539–542. <https://doi.org/10.1093/sysbio/sys029>
- Shan RH (1943) Umbelliferae novae sinicae. Sin. Special Bull 14: 111–114.
- Shan RH, Sheh ML (1992) Umbelliferae. Flora Reipublicae Popularis Sinicae, vol. 55. Science Press, Beijing.
- She ML, Phillippe LR (2005) *Sanicula* Linnaeus In: Flora of China, vol. 14. Science Press, Beijing & Missouri Botanical Garden Press, St. Louis, 19–24.
- She ML, Pu FD (1997) A new species of *Notopterygium* de Bois. from China. Journal of Plant Resources and Environment 6(2): 41–42.
- She ML, Watson MF (2005a) *Notopterygium* H. de Boissieu In: Flora of China, vol. 14. Science Press, Beijing & Missouri Botanical Garden Press, St. Louis, 53–55.
- She ML, Watson MF (2005b) *Haplosphaera* Hand.-Mazz. In: Flora of China, vol. 14. Science Press, Beijing & Missouri Botanical Garden Press, St. Louis, 152–153.
- Stamatakis A (2014) RAxML version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics 30(9): 1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>
- Tan JB, Jia SB, He XJ, Ma XG (2020) Accommodating *Haplosphaera* in *Hansenia* (Apiaceae) based on morphological and molecular evidence. Phytotaxa 464(3): 207–216. <https://doi.org/10.11646/phytotaxa.464.3.2>
- Turczaninow NS (1844) Flora baicalensi-dahurica seu descriptio plantarum in regionibus ciset transbaicalensibus atque in Dahuria sponte nascentium, Ordo XXXIV, Umbelliferae. Naturalistes Moscou 17: 707–754.

- Wang YP, Pu FD, Wang PL, He XJ (1996) Studies on the systematics of the chinese endemic genus *Notopterygium*. *Acta Botanica Yunnanica* 18(4): 424–430.
- Wei J, Gao YZ, Zhou J, Liu ZW (2019) Collection and sorting of medicinal plants in Chinese Apiaceae (Umbelliferae). *China Journal of Chinese Materia Medica* 44(24): 5329–5335.
- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) PCR Protocols: A Guide to Methods and Applications. Academic Press Inc, San Diego, California, 315–322. <https://doi.org/10.1016/B978-0-12-372180-8.50042-1>
- Winter PJD, Magee AR, Phephu N, Tilney PM, Downie SR, van Wyk B-E (2008) A new generic classification for African peucedanoid species (Apiaceae). *Taxon* 57(2): 347–364. <https://doi.org/10.2307/25066009>
- Wolff H (1930) Umbelliferae asiaticae novae relictæ (III). *Repertorium novarum specierum regni vegetabilis* 27(16–25): 301–335. <https://doi.org/10.1002/fedr.4870271612>
- Zhang D, Gao F, Jakovlić I, Zou H, Zhang J, Li WX, Wang GT (2020) PhyloSuite: An integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. *Molecular Ecology Resources* 20(1): 348–355. <https://doi.org/10.1111/1755-0998.13096>

Supplementary material I

Figure S1, S2

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Data type: Docx file.

Explanation note: **Figure S1.** Fruit of *Hansenia trifoliolata*. **Figure S2.** Isotype of *Hansenia trifoliolata* and paratype of *H. trifoliolata*.

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